



Determination of Acrylamide in Branded and Non-branded Potato Chips by using High Performance Liquid Chromatography

*Naseem Zahra, Qurat-ul-Ain Syed, Imran Kalim, Zohaib Khurshid,
Ijaz Ahmad and Muhammad Khalid Saeed

Food and Biotechnology Research Centre, PCSIR Laboratories Complex, Ferozepur Road, Lahore-54600, Pakistan.

Corresponding Author Email: naseem.zahra1981@gmail.com

Received 05 December 2017, Revised 21 May 2018, Accepted 24 May 2018

Abstract

Acrylamide, a starchy food process contaminant is carcinogenic, neurotoxic and genotoxic substance. It is formed at temperatures of 100°C- 220°C. The chips are very popular food items especially among children. Deep Frying of chips may cause production of acrylamide. The presence of acrylamide in chips may result in severe health issues in consumers especially in children. By keeping this aspect in view, the present study was conducted to determine acrylamide in different potato chips of branded and non-branded quality. The highest amount of acrylamide was determined in chips sample K obtained from local store (nonbranded) in Lahore i.e. 2649.80 µg/kg and the lowest concentration of acrylamide was detected in sample taken from local vendor i.e. 390.25 µg/kg. The astounding results were obtained as branded chips have considerable amount of acrylamide. Formation of acrylamide in fried chips is matter of concern as potato chips are widely consumed in Pakistan especially by children. It is dire need to have awareness of this probable carcinogen as consumers and industries still have no knowledge about this chemical's carcinogenicity. Therefore, without delay work should be started to mitigate the acrylamide levels in food commodities during processing.

Keywords: Acrylamide, Potato, Chips, HPLC, Retention time.

Introduction

Acrylamide (AA) or 2-propenamide formed in a broad range of food products (oven cooked and fried foods) was discovered for the first time in 2002 [1, 2]. The unearthing of acrylamide in foods raised great concerns for the reason that acrylamide has been classified a potent carcinogen [3]. Acrylamide in food is produced as a result of the Maillard reaction among asparagine and reducing sugars. Thus, the amount of acrylamide depends on the concentration of these antecedents in food samples, alongwith different processing conditions during food preparation and storage [4]. It is concluded that acrylamide is formed as a result of asparagine degradation by carbonyl (from glucose and fructose source) reaction [5, 6]. There

are many factors which may affect formation of acrylamide in potato and cereals products i.e. variety of raw materials, harvesting time, storage condition and fertilization [7-10]. The limiting factor in acrylamide formation process are reducing sugars, so it is better option to control reducing sugars and asparagine to reduce acrylamide concentration [11].

There are different procedures for the determination of acrylamide in food items i.e. liquid chromatography [12], gas chromatography [13] and mass spectrometry coupled with gas chromatography [14] or liquid chromatography [15]. HPLC is very good, low cost and sensitive

technique to determine acrylamide concentration in potato chips by acetone extraction. It was found that in french fries acrylamide concentration levels increased as the oven temperature increased from 100-220°C and achieved a maximum level of 5000µg/kg [16]. Acrylamide formation begins to start at 100°C and increases upto 220°C in processes like frying, roasting and baking. It is estimated by conducting risk assessment that daily intake of acrylamide is 0.2-0.8 mg/kg body weight [17].

According to European Union the acrylamide concentration was found high in processed foods like fried chips, potato chips, crispy bread, biscuits, breads and cereals [18]. Acrylamide in highest concentrations were detected in coffee (instant) and coffee substitutes. Although some food companies have taken necessary actions to reduce the amount of acrylamide in different products but overall scenario shows that more drastic efforts are needed to deal with this serious issue concerning public health [19]. The current study is conducted to determine acrylamide presence and concentrations in potato chips of branded, local bakery and street vendor quality.

Materials and Methods

Reagents

All the chemicals and reagents used in the present study were purchased from Merck (Darmstadt, Germany).

Samples collection

12 chips samples were purchased from local stores and bakeries. 8 samples were of two popular brands in Lahore, Pakistan while 4 samples were collected from local vendor and bakeries.

Acrylamide extraction

Acrylamide extraction was done by following method described [20] by Koshnam et al., 2010. Sample was crushed using mortar and pestle to homogenize the sample. 10 g sample was weighed using analytical weighing apparatus. 25ml

of n-Hexane was added in flask containing sample to de-fat the sample by putting it on wrist action shaker for 10 min. This step was repeated for thorough de- fattening. Hexane was decanted and sample was dried by using hot plate on moderate temperature. For extraction of acrylamide, 50ml of acetone and 100µl of distilled water in the sample were added. Flask was placed on water bath preset at 40°C for 20 min. After 20 min, acetone was filtered and half of it was evaporated by putting it on hot plate again. Suspended the residue in 5 ml of distilled water and filtered it with 0.25 micron syringe filter for further HPLC analysis.

Preparation of acrylamide standard solution

Standard of Acrylamide (Sigma Aldrich) was purchased to carry out HPLC analysis. Stock solution for standard was prepared by weighing 10.70 mg acrylamide precisely and dissolving in 50 ml distilled water which makes concentration of sample 0.214 mg/ml or 214 ppm. Stock solution was further diluted to construct calibration graph for quantification and kept away from direct sunlight and stored under refrigeration.

HPLC analysis

Samples were analyzed by using HPLC Perkin-Elmer 200 Series with C-18 Column having 250mm length, 4.6 mm diameter and 25 micron particle size. Column oven temperature was set at 40°C and flow rate was maintained at 1ml/min. The analysis was performed at 202nm with a UV detector. First of all, standard solution was run and then samples were run and their retention time was compared with standard solution.

Results and Discussion

Acrylamide is not a matter that is added to food entities but it is formed in food during heat processing. Present research was designed to determine acrylamide in potato chips of branded and non branded type alongwith local shops of Lahore in October 2015-March 2016. Total 12 samples were checked in triplicate for the presence of acrylamide concentration.

High performance liquid chromatographic technique was used to determine acrylamide in potato chips due to its effectiveness and reliable results. The mobile phases used were 80% acetonitrile and 20% distilled water. Standard acrylamide was determined with characteristic peak at 3.04 minutes of retention time. Limit of detection (LOD) and limit of quantification (LOQ) for the present method was 2.46 and 3.14 ng g⁻¹, respectively [20].

In the study conducted by Geng et al., to analyze acrylamide in potato chips, LOD and LOQ values were 15 and 50 µg/kg correspondingly [21]. Hariri et al., 2015 looked into corn and potato chips for the detection of acrylamide and heavy metals. It was estimated that potato chips contained 25% more acrylamide than corn chips. Shocking fact was that acrylamide intake was 7-40 times greater than that of risk intake for causing cancer set by World Health Organization [22]. The results of acrylamide in potato chips and Average, maximum and minimum are given in Table 1.

Table 1. Results of acrylamide determination in potato chips by HPLC.

Sample No.	Sample Code	Mean Retention Time (minutes)	Mean Quantity (µg/kg) ± S.D.
S1	Branded A	3.37	905.00 ± 1.000
S2	Branded B	2.97	748.53 ± 1.000
S3	Branded C	2.78	821.36 ± 1.000
S4	Branded D	2.79	689.62 ± 0.510
S5	Branded E	3.22	547.04 ± 0.031
S6	Branded F (Imported)	2.82	1002.86 ± 1.010
S7	Branded G	2.79	989.45 ± 0.810
S8	Branded H (Imported)	3.18	780.61 ± 0.315
S9	Local I- Non branded (Vendor)	3.20	390.25 ± 0.510
S10	Local J- Non branded	3.20	871.89 ± 0.210
S11	Local K- Non branded	3.17	2649.80 ± 1.100
S12	Local L- Non branded	2.97	458.74 ± 0.510

D.D. = Standard deviation of triplicate analysis

In an investigation; acrylamide concentration in food products purchased from Latvian local market. There were 8 sets of samples which consisted of rye based breads, wheat based breads, potato chips, coffee etc. Ultra high performance liquid chromatography attached with tandem mass spectrometric detector was used in this study [23]. Results had shown higher acrylamide concentration in foods which are processed e.g. potato chips and coffee as compared to unprocessed foods. Range of acrylamide concentration was from 564 µg/kg to 2790 µg/kg while on the other hand not any single bread have shown acrylamide presence higher than allowable limits except for the breads which contain seeds. More significantly, presence of fruits and vegetables in breads has an escalating outcome on acrylamide concentration.

In another study; the fried chips of brand kurkure was found to have the maximum acrylamide concentration when compared with other bakery and puff food stuff. Kurkure's acrylamide concentration was measured to be the highest due to various factors such as compositional difference and degree of frying when compared with other processed food products [24].

In current study, the highest amount of acrylamide was determined in chips sample K obtained from local store of Lahore i.e. 2649.80 µg/kg and the lowest concentration of acrylamide was detected in sample I taken from local vendor i.e. 390.25 µg/kg while the branded samples also have considerable amounts of acrylamide also; which is notable (Fig. 1).

Similar studies were conducted to obtain the data on acrylamide in various food commodities from different companies and provided an overview regarding acrylamide situation and health hazards of branded cakes, biscuits and potato chips in Pakistan [25]. The concentrations of acrylamide in biscuits were found in the range of 52.3 ± 0.70 to 507.5 ± 1.5 µg/kg; while in cakes it was less while in potato chips the concentrations were between 27.1 ± 0.65 to 1323.0 ± 3.0 µg/kg.

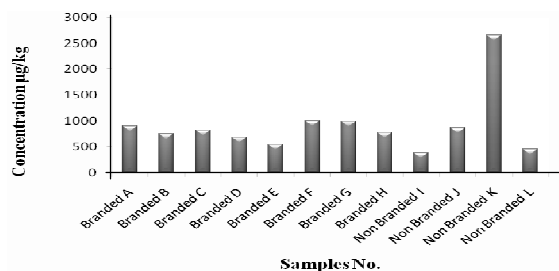


Figure 1. Concentration of acrylamide in various chips samples

Acrylamide, a potent carcinogenic compound is produced in baked and fried food commodities due to Maillard reaction. In study conducted on French fries collected from Faisalabad and Lahore, it was found that acrylamide contents were higher (beyond permissible daily intake) in homemade and local vendor samples [26]. It is in contrast with present study in which; it is worth noted that acrylamide was in low concentration in sample obtained from local vendor while chips samples taken from mega store and branded samples have highest amounts of acrylamide.

The study conducted on different products showed that chocolate (11.2%), bread (23.5%) while French fries (29.9%) are the major sources of dietetic acrylamide. Food commodities used among the three main meals of the day (so called snack type foods) added the most to the intake (42.2%). So, the French fries may contain highest amounts of acrylamide which can harm human health [27].

The presence of acrylamide in potato chips can be linked with oil quality and processing time. Furthermore, different pre-treatments like freezing, frying and both can affect acrylamide formation and its reduction as well. Different ingredients may also enhance acrylamide concentrations. Such highest concentrations of acrylamide may impact health very badly as it found to be very prone to cancer.

Conclusion

The purpose of the present work was to search out the presence of acrylamide in potato chips of branded and non branded samples. It is concluded that acrylamide might be produced in

the fried chips due to the chemical reaction started during high temperature cooking. Although differences are present in the amount of acrylamide among all samples but all have notable amount of acrylamide. Due to omnipresent nature of acrylamide it is necessary to take mitigation steps to avoid health hazardous issues due to acrylamide.

References

1. M. M. Nassar and Y. H. Magdy, *Chem. Eng. J.*, 66 (1997) 223.
[doi.org/10.1016/S1385-8947\(96\)03193-2](https://doi.org/10.1016/S1385-8947(96)03193-2)
2. V. Garg, R. Gupta, A.B. Yadav and R. Kumar, *Biores. Tech.*, 89 (2003) 121.
[doi.org/10.1016/S0960-8524\(03\)00058-0](https://doi.org/10.1016/S0960-8524(03)00058-0)
3. S. Saiful Azhar, A. Abdul Ghaniey Liew, D. Suhardy, K. Farizul Hafiz and M. I. Hatim, *Amer. J. Appl. Sci.*, 2 (2005) 1.
<https://doi.org/10.1051/mateconf/201710306015>
4. A. Bhatnagar, E. Kumar, A. Minocha, B-H Jeon, H. Song and Y-C Seo, *Sep. Sci. Tech.*, 44 (2009) 316.
<https://doi.org/10.1080/01496390802437461>
5. R. Sivaraj C. Namasivayam and K. Kadirvelu, *Waste Manage.*, 21 (2001) 105.
[https://doi.org/10.1016/S0956-053X\(00\)00076-3](https://doi.org/10.1016/S0956-053X(00)00076-3)
6. O. Amuda O, Alade A. *Desalination.* 196 (2006) 22.
[doi: 10.1016/j.desal.2005.10.039](https://doi.org/10.1016/j.desal.2005.10.039)
7. M. Yazdanbakhsh, H. Tavakkoli and S. M. Hosseini, *Desalination*, 281 (2011) 388.
<https://doi.org/10.1016/j.desal.2011.08.020>
8. T. Tabari, H. Tavakkoli, P. Zargarani and D. Beiknejad, *South Afri. J. Chem.*, 65 (2015) 239.
<https://www.ajol.info/index.php/sajc/article/view/123859/113422>
9. P. Brown, I. A. Jefcoat, D. Parrish, S. Gill and E. Graham, *Adv. Env. Res.*, 4 (2000) 19.
[https://doi.org/10.1016/S1093-0191\(00\)00004-6](https://doi.org/10.1016/S1093-0191(00)00004-6)
10. V. C. Taty-Costodes, H. Fauduet, C. Porte and A. Delacroix, *J. Hazard. Mater.*, 105 (2003) 121.
<https://doi.org/10.1016/j.jhazmat.2003.07.009>
11. K. Singh, M. Talat and S. Hasan, *Biores. Tech.*, 97 (2006) 2124.

12. <https://doi.org/10.1016/j.biortech.2005.09.016>
S. Chatterjee, S. Chatterjee, B. P. Chatterjee, A. R. Das and A. K. Guha, *J. Colloid Inter Sci.*, 288 (2005) 30.
<https://doi.org/10.1016/j.jcis.2005.02.055>
13. K. Vijayaraghavan, K. Palanivelu and M. Velan, *Biores. Tech.*, 97 (2006) 1411.
<https://doi.org/10.1016/j.biortech.2005.07.001>
14. D-K. Kweon, J-K. Choi, E-K. Kim and S-T. Lim, *Carb. Polymer*, 46 (2001) 171.
[https://doi.org/10.1016/S0144-8617\(00\)00300-3](https://doi.org/10.1016/S0144-8617(00)00300-3)
15. K. N. Ghimire, K. Inoue, T. Miyajima, K. Yoshizuka and T. Shoji, *Chitin and Chitosan Res.*, 7 (2001) 61.
<https://ci.nii.ac.jp/naid/10028235276/>
16. U. Kumar and M. Bandyopadhyay, *Biores. Tech.*, 97 (2006) 104.
<https://doi.org/10.1016/j.biortech.2005.02.027>
17. R. Gong, X. Zhang, H. Liu, Y. Sun and B. Liu, *Bioresource Tech*, 98 (2007) 1319.
<https://doi.org/10.1016/j.biortech.2006.04.034>
18. M. Husseien, A. Amer, A. El-Maghraby and N. A. Taha, *J. Appl. Sci. Res.*, 3 (2007) 1352.
19. R. P. Dhakal, K. N. Ghimire, K. Inoue, M. Yano and K. Makino, *Sep. Puri. Tech.*, 42 (2005) 219.
<https://doi.org/10.1016/j.seppur.2004.07.016>
20. N. Pramanpol and N. Nitayapat, *Kasetsart J.*, 40 (2006) 192.
https://www.researchgate.net/profile/Nuttakan_Nitayapat/publication/228969881_Adsorption_of_reactive_dye_by_eggshell_and_its_membrane/links/09e4150fe079b58101000000.pdf
21. W-T. Tsai, H-R. Chen, K-C. Kuo, C-Y. Lai, T-C. Suand and Y-M. Chang, *J. Env. Eng. Manage. J.*, 19 (2009) 165.
http://ser.cienve.org.tw/download/19-3/jeeam19-3_165-172.pdf
22. K. V. Kumar, *Dye Pigments*, 74 (2007) 595.
<https://doi.org/10.1016/j.dyepig.2006.03.026>
23. S.R. Gillani, , Z. Mahmood, M. Imran, A. Saeed and S. Hamid, *J. Chem. Soc. Pak.*, 33 (2011) 364.
<https://www.jcsp.org.pk/ViewByVolume.aspx?v=106&i=VOLUME%2033,%20NO3,%20JUN%202011>
24. C. Tejada, *Ingenieria y Universidad*, 19 (2015) 37.
<http://dx.doi.org/10.1114/javeriana.iyu19-2.kamr>
25. National Center for Biotechnology Information. PubChem Compound Database; CID=11048,
<https://pubchem.ncbi.nlm.nih.gov/compound/11048> (accessed Mar. 12, 2018).
26. K. Ravikumar, B. Deebika and K. Balu, *J. Hazard. Mater.*, 122 (2005) 75.
<https://doi.org/10.1016/j.jhazmat.2005.03.008>
27. E. El-Katori, A. Fouda and A. Al-Sarawy, *Indian J. Chem. Tech.*, 18 (2011) 319.
<http://nopr.niscair.res.in/bitstream/123456789/12664/1/IJCT%2018%284%29%20319-326.pdf>
28. S. Allen and B. Koumanova, *J. Uni. Chem. Tech. Metallurgy*, 40 (2005) 175.
<http://dl.uctm.edu/journal/node/j2005-3/Review.pdf>
29. D. Gialamoudis, M. Mitrakas and M. Liakopoulou-Kyriakides M. *J. Hazard. Mater.*, 182 (2010) 672.
<https://doi.org/10.1016/j.jhazmat.2010.06.084>
30. I. Langmuir. *J. Amer. Chem. Soc.*, 38 (1916) 2221.
pubs.acs.org/doi/pdf/10.1021/ja02268a002
31. O. Abdelwahab, *Desalination*, 222 (2008) 357.
<https://doi.org/10.1016/j.jcis.2005.02.055>